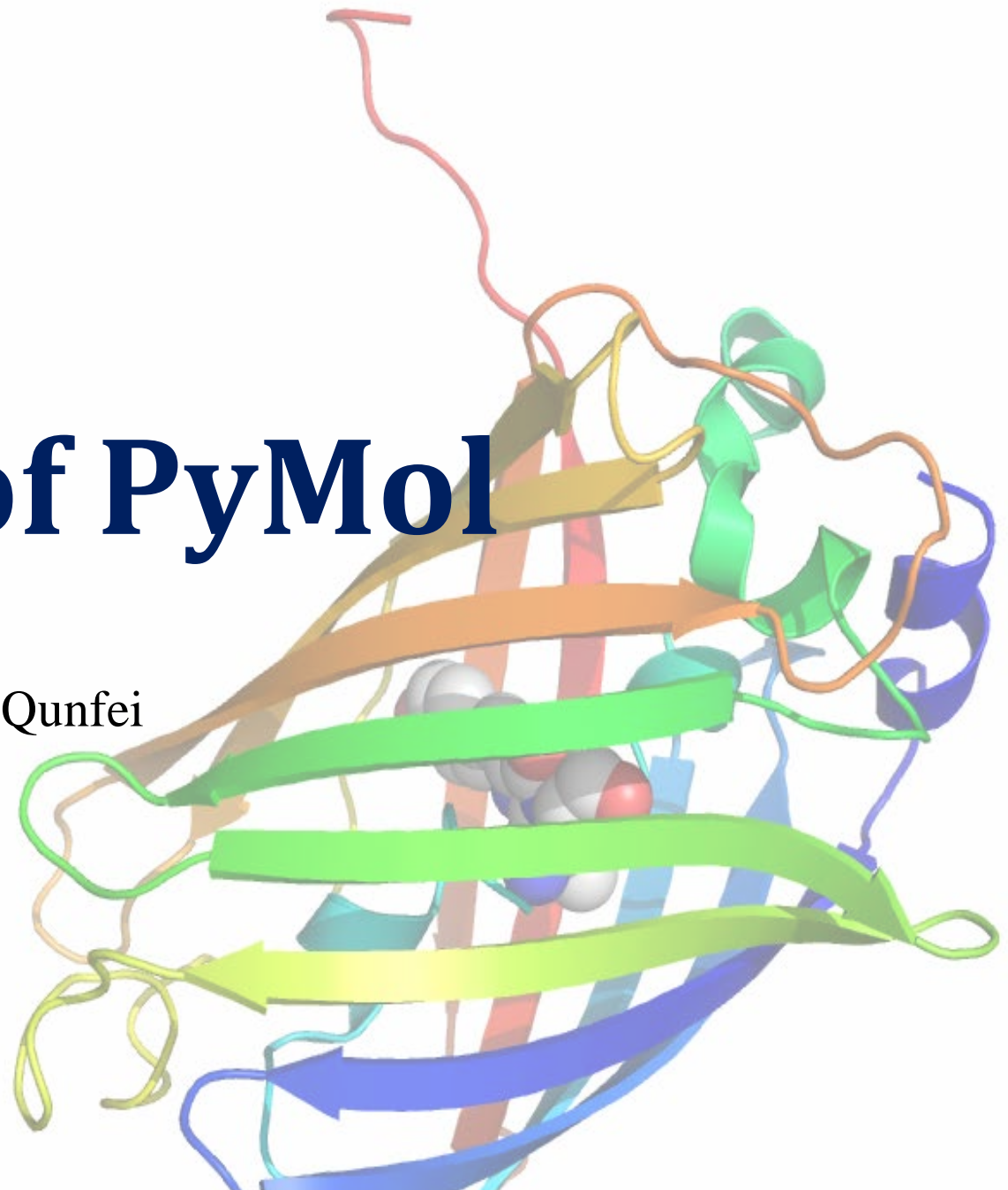


Learning notes of PyMol

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Content

- **What is PyMol?**
- **What PyMol looks like?**
- **Why PyMol can draw amazing pictures?**
- **What can we do by using PyMol?**
 - Measurement
 - Structure alignment
 - Electrostatics surface
 - Highlight the ligand and the interactions
 - Electron density map
 - Making movies



What is PyMol?

A Powerful Desktop Molecular Graphical Tool which is also making progress to become a research tool!!

Why we choose PyMol?

Visualization is essential to understanding structural biology.

- (1) Visualize multiple conformations of a single structure;
- (2) Interface with external programs;
- (3) Provide professional strength graphics under both Windows and Unix;
- (4) Prepare publication quality images;
- (5) Fit into a tight budget.

What's the differences between PyMol and Swiss-pdbViewer?

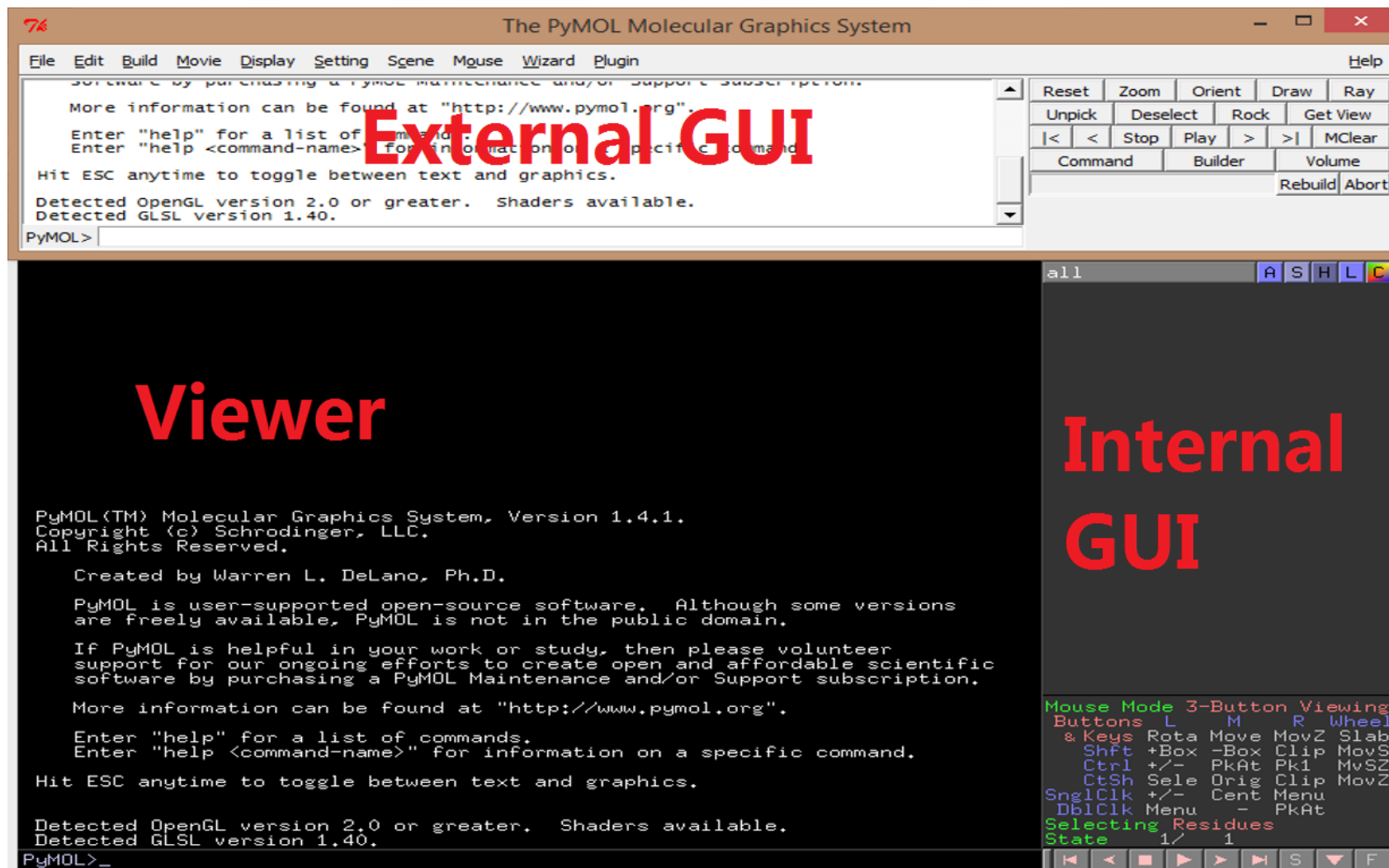
I think...

Swiss-pdbViewer is more likely to be a tool for scientific research,
Pymol is more likely to be a tool for scientific results presentation.

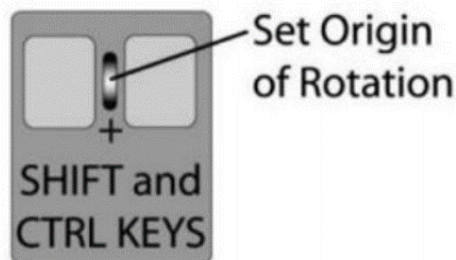
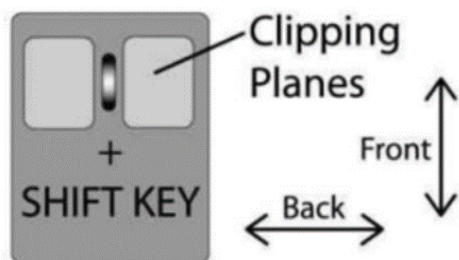
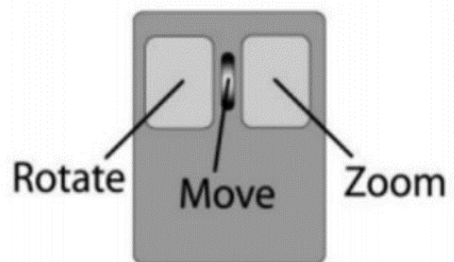
What PyMol looks like?

PyMol's Interface

GUI:
Graphical User
Interface



Viewer Window & Mouse Using



PyMOL Viewer

Display>sequence
Your macromolecule's sequence will be shown

Names Panel
A: Action; S: Show; H: Hide; L: Label; C:

Mouse Matrix
just see here if you foget how to use your mouse

Movie Controls

Command Line

```
PyMOL>_
```

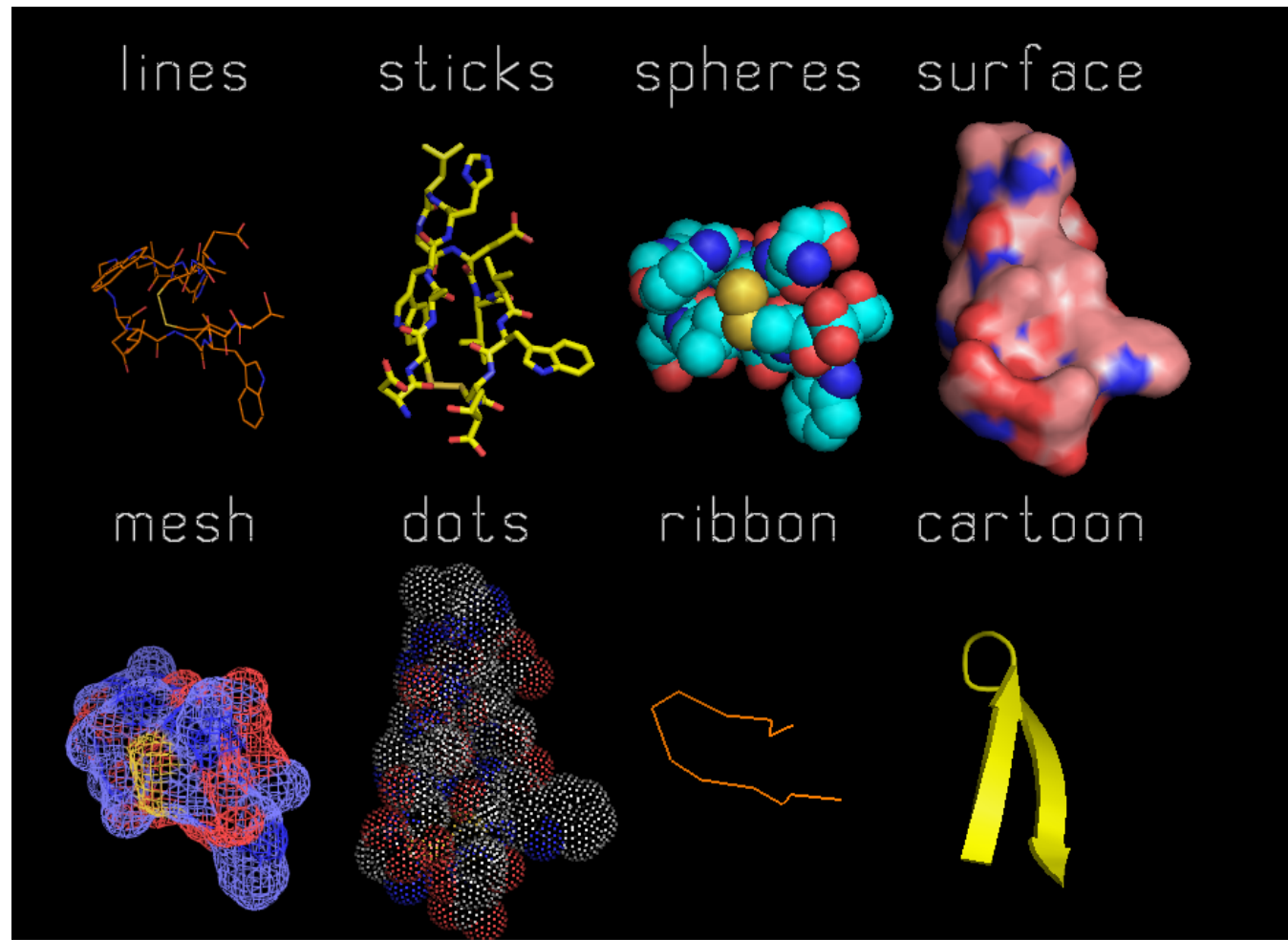
PyMOL Viewer interface showing a protein structure, command line, names panel, mouse matrix, and movie controls.

Why PyMol can draw amazing pictures?

Before starting learning PyMol, you can click:

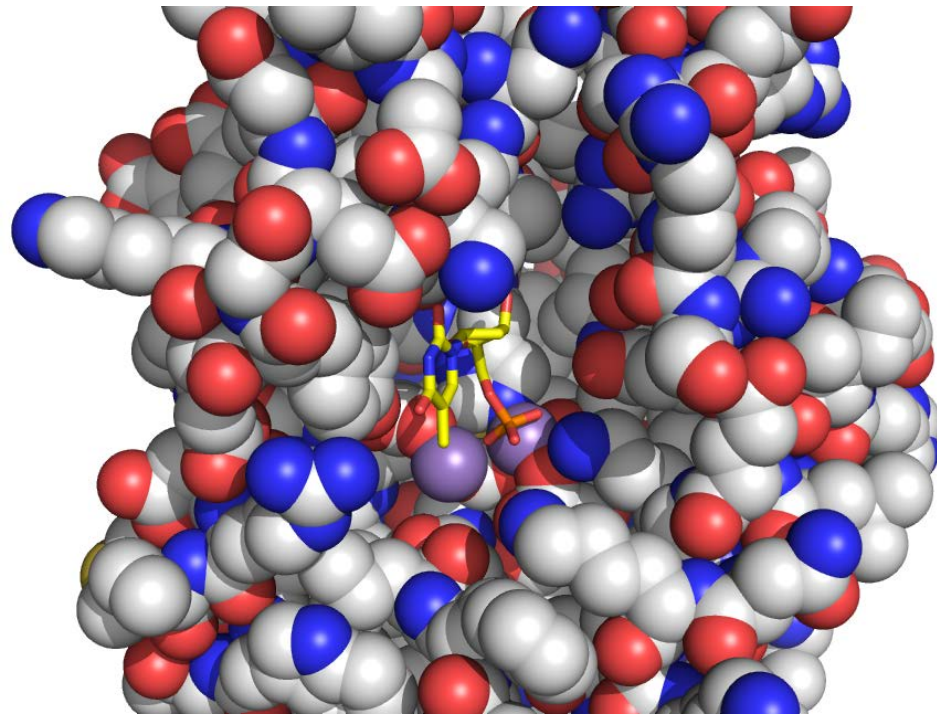
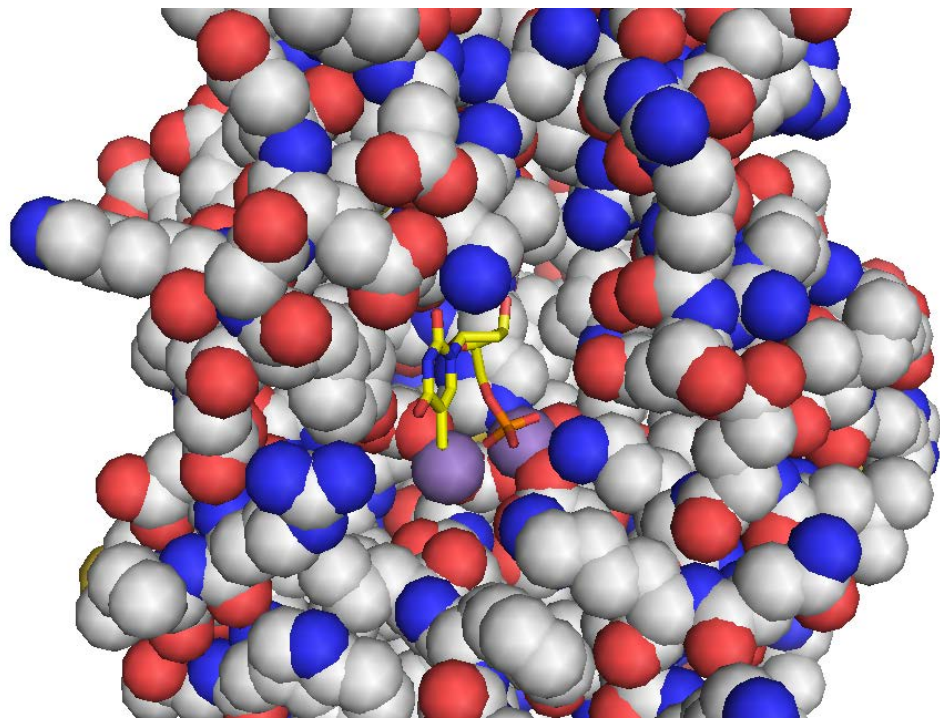
>Wizard>Demo>...

To see what PyMol can do for you.

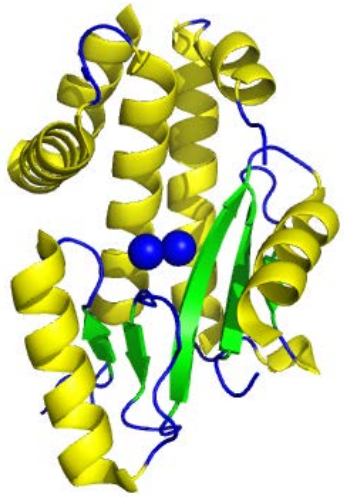


• **Fancy ray**

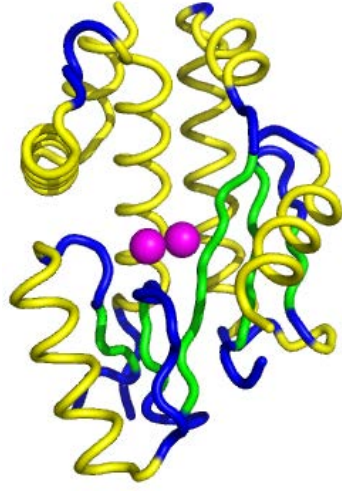
>ray



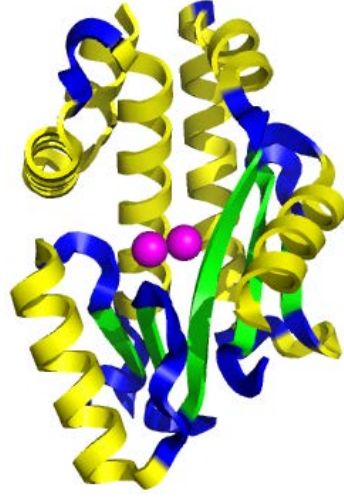
- **Fancy cartoon**



Automatic



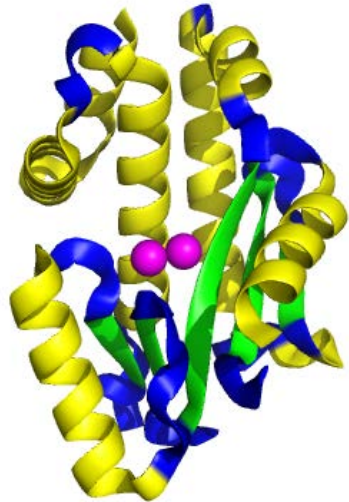
Tube



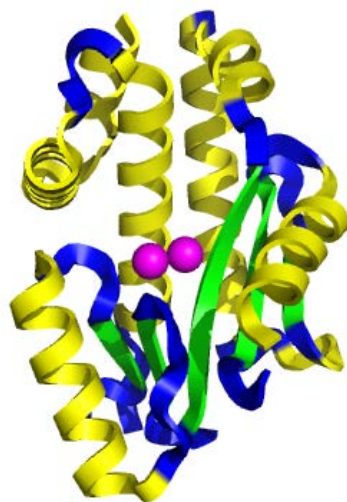
Arrow



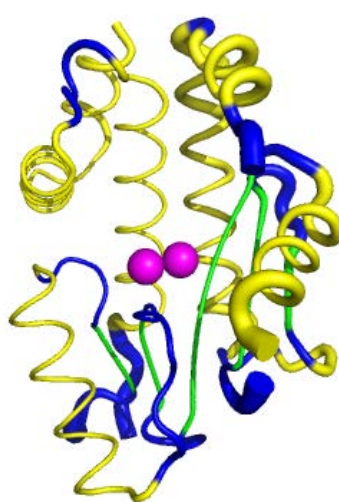
Loop



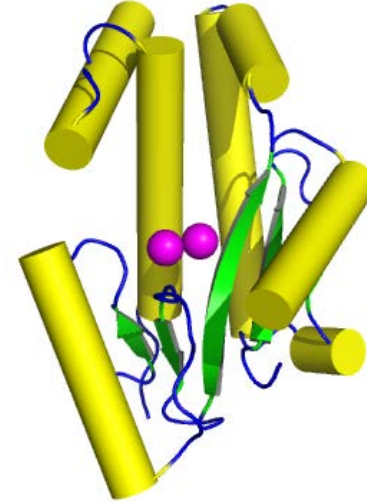
Oval



Rectangle



putty



Setting > Cartoon >...

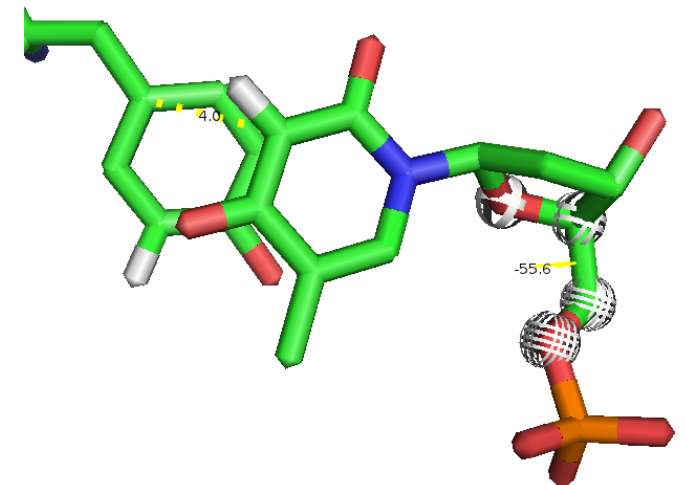
You can define the cartoon as what you like.

How to use PyMol ?

Measurement-Advanced Mouse operation

- Measure the distance between two atoms:
Mouse:
Ctrl+R; %select the first atom;
Ctrl+M; %select the second atom;
Command:
>distance/dist
- Measure the angle between three atoms:
Mouse:
Ctrl+R; %select the first atom;
Ctrl+M; %select the second atom;
Ctrl+M; %select the third atom;
Command:
>angle/ang

- Measure the dihedral formed by four atoms:
Mouse:
Ctrl+R; %select the first atom;
Ctrl+M; %select the second atom;
Ctrl+M; %select the third atom;
Ctrl+M; %select the fourth atom;
Command:
>dihedral/dih



Structure alignment

Fit by protein sequence

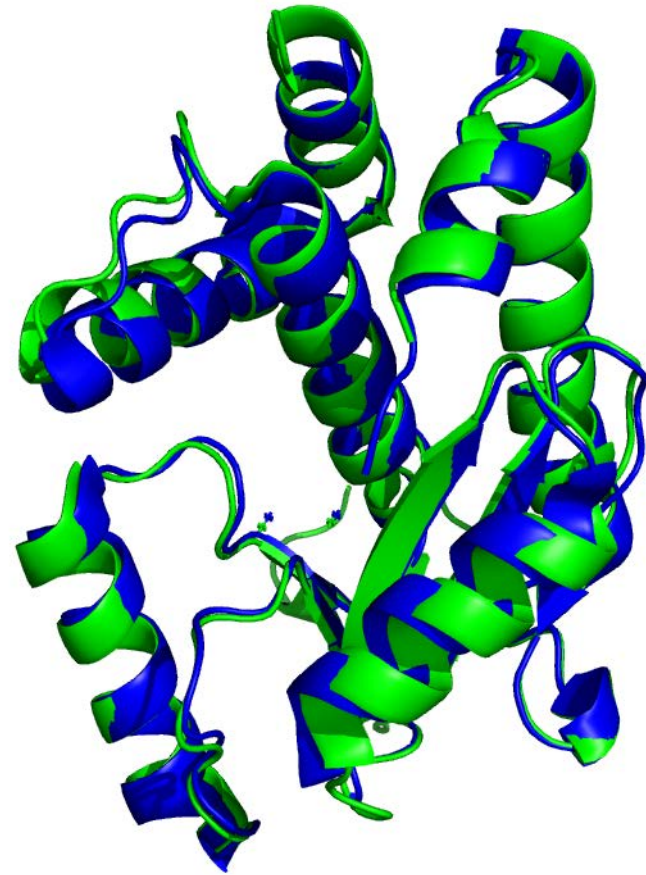
- Mouse:
- First molecule>A>align>to molecule Second molecule;
- Command:
- >Align (A and name ca), (B and name ca)

Fit by overall structure

- Command:
- >cealign A, B

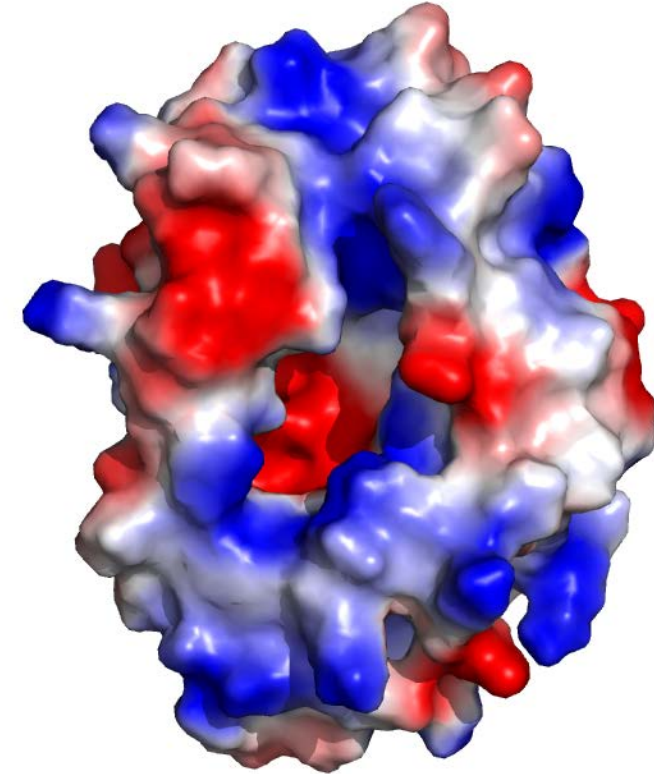
Fit by atom pair

- Mouse:
- Wizard>Pair Fitting



Show the simple electrostatics surface

- Your protein >A> generate >vacuum electrostatics>protein contact potential (local)
- **Note:** Due to short cutoffs, truncations, and lack of solvent “screening”, these computed potentials are only qualitatively useful.
- You can install the package “APBS” for accurate calculation.



How about doing something that Swiss-pdbViewer can not do?

Before that...

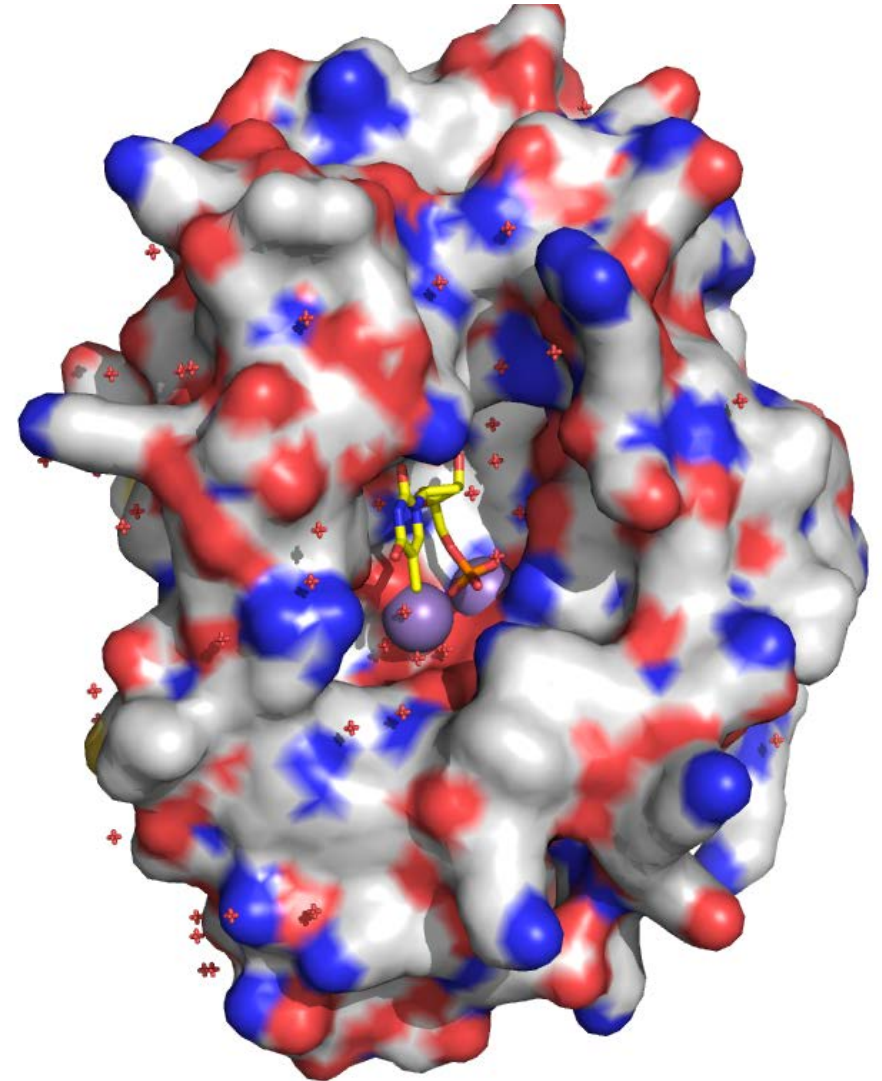
The example protein will be used: PA polymerase of Influenza virus.

PDB code: 4AVL

Function: “Cap snatching” of host pre-mRNA

Active site: 3000 Å²; Metal coordination; highly conserved

Ligand: TMP



Highlight the ligand and the interactions

1. Draw the scaffold;

```
4AVL>H>everything;
```

```
4AVL>S>cartoon;
```

```
4AVL>C>by ss;
```

2. Draw the ligand;

```
Display>Sequence;
```

```
# Select TMP with mouse;
```

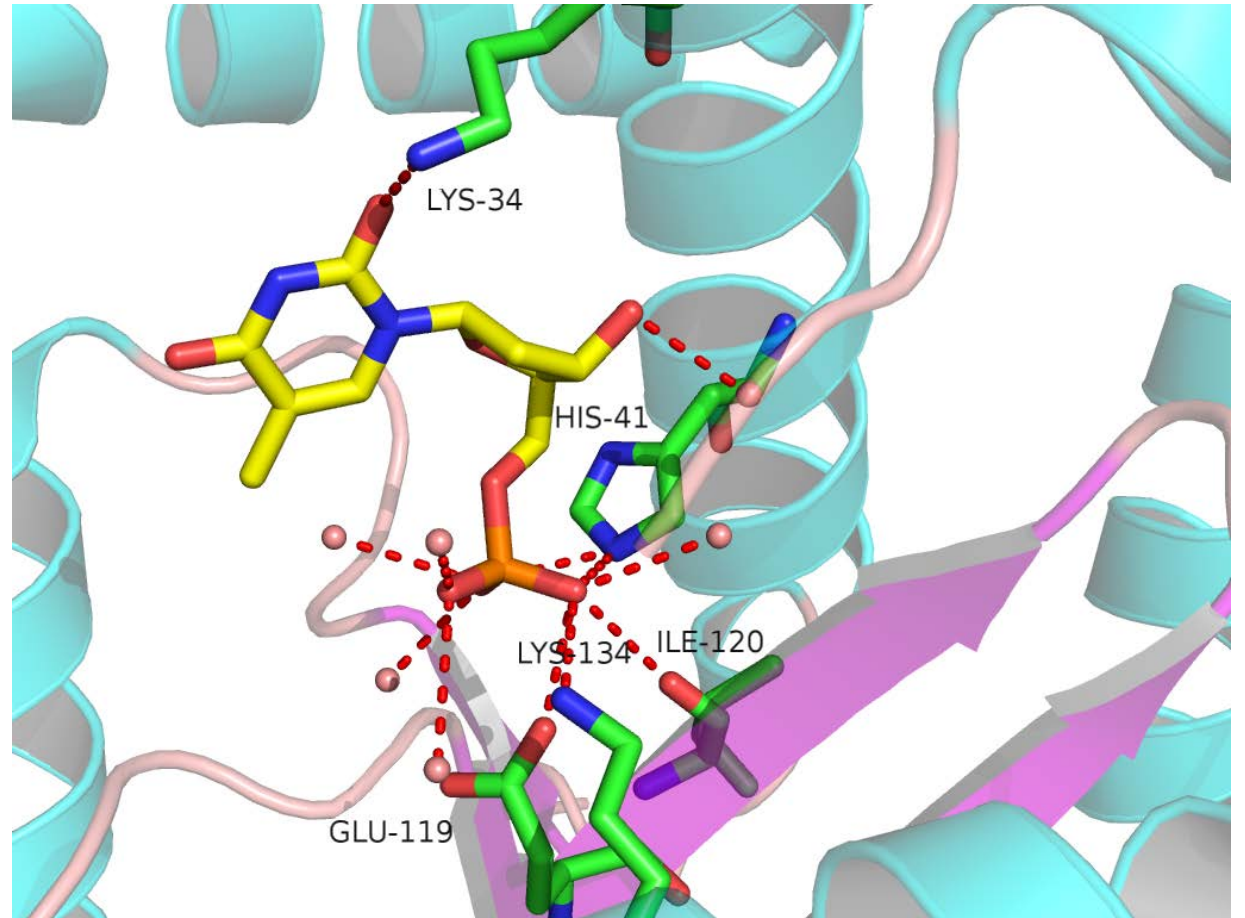
```
(sele)>A>find>polar contacts>to other atoms in  
the object;
```

```
(sele)>A>copy to object;
```

```
obj01>A>rename object: lig;
```

```
lig>S>sticks;
```

```
lig>C>by element>yellow...
```



3. Draw the interactions

```
Lig>A>orient;
```

```
4AVL>show lines;
```

```
# Find the residues that contact with the lig, select them with mouse;
```

```
(sele)>A>copy to object;
```

```
obj02>A>rename object: resi_cont;
```

```
resi_cont>S>sticks;
```

```
resi_cont>C>by element>blue...
```

```
4AVL>H>lines;
```

```
4AVL>S>nb_sphere;
```

```
# Find the HOHs that contact with the lig, select them with mouse;
```

```
(sele)>A>copy to object;
```

```
obj03>A>rename object: HOH_cont;
```

```
4AVL>H>nb_sphere;
```

```
HOH_cont>S>nb_sphere;
```

```
HOH_cont>C>red;
```

4. Draw the metal ions;

```
# Find the MN in the sequence list, select them with mouse;
```

```
(sele)>A>copy to object;
```

```
obj04>A>rename object: metal
```

5. Optimize the picture

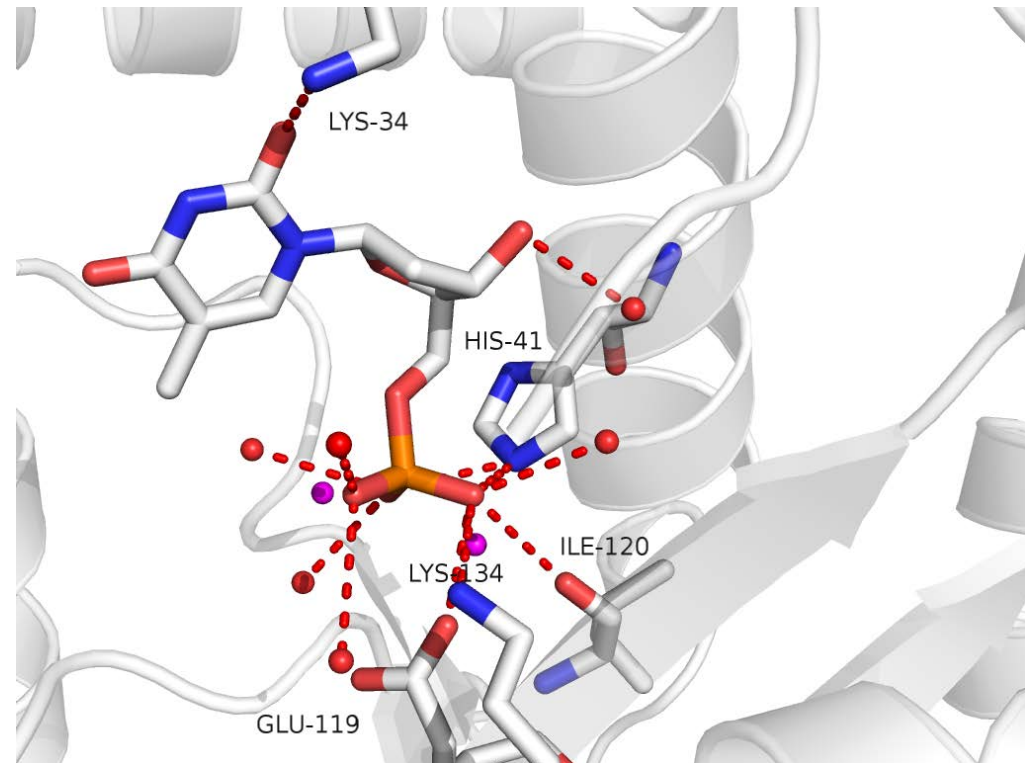
```
Display>background>white;
```

```
Setting>cartoon>fancy helices;
```

```
Setting>cartoon>highlight colors;
```

```
Setting>transparency>caroon>50%;
```

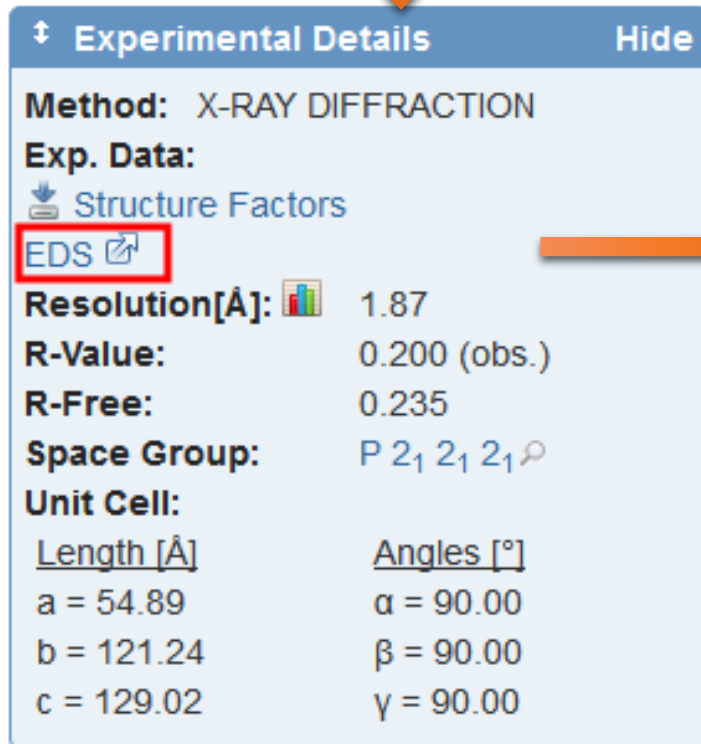
```
Command: ray 1024, 768
```



Electron Density Map

First Step: Download an electron density map

- >Go to the PDB Database
- >input your PDB code (eg. 4AVL)
- >find this window



Experimental Details

Method: X-RAY DIFFRACTION

Exp. Data:

Structure Factors

EDS

Resolution[Å]: 1.87

R-Value: 0.200 (obs.)

R-Free: 0.235

Space Group: $P 2_1 2_1 2_1$

Unit Cell:

Length [Å]	Angles [°]
a = 54.89	$\alpha = 90.00$
b = 121.24	$\beta = 90.00$
c = 129.02	$\gamma = 90.00$



Download

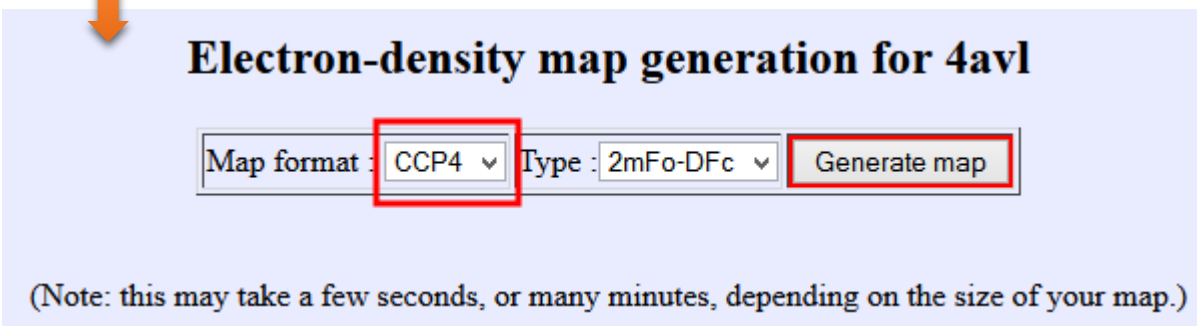
[Coordinates](#)

[Maps](#)

[Statistics](#)

[All files \(.tar.gz\)](#)

- >Download your map file (eg. 4avl.ccp4)
 - >Uncompressed it;
 - >Rename it: 2biw.map.ccp4
- (You will find it make the work convenient!!)



Electron-density map generation for 4avl

Map format : CCP4 Type : 2mFo-DFc Generate map

(Note: this may take a few seconds, or many minutes, depending on the size of your map.)

Second step: Display electron density map

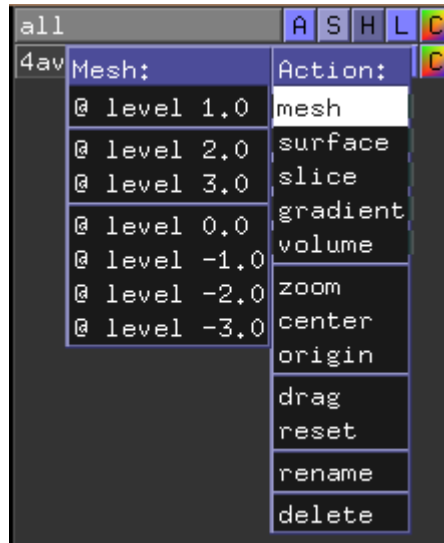
> Load your map file;

☺ Do not panic if your screen still appears black at this point! This is normal.

```
4avl>mesh>@ level 1.0/2.0/3.0
```

☺ Do not panic if you find a lot of white worms! This is normal.

(The increase of electron density map means the decreasing resolution of the map at this site)



Third step: Add your PDB file

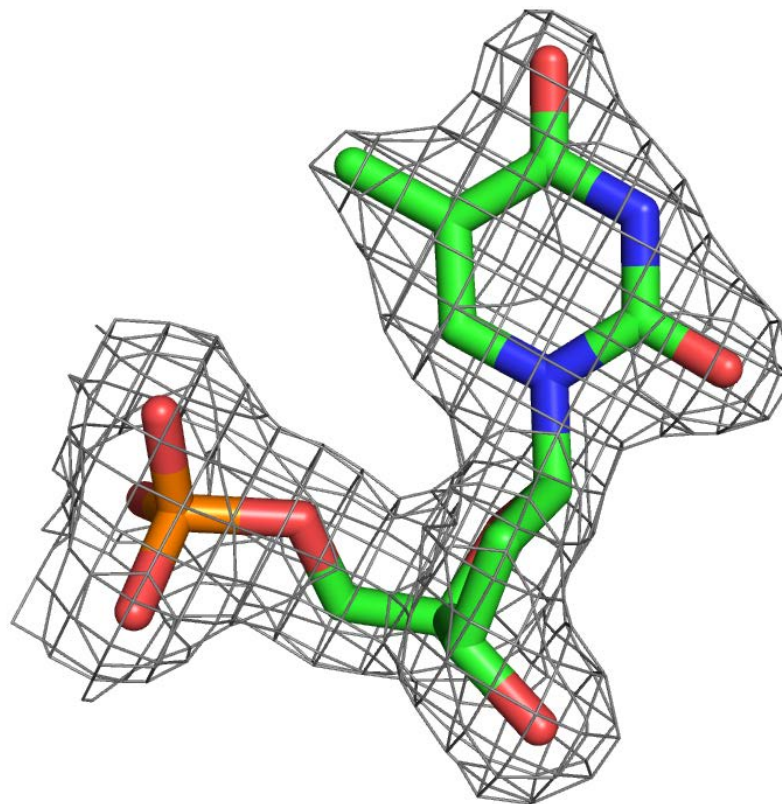
```
>load 4avl.pdb
```

Fourth step: Display electron density map for what you want

```
Display>Sequence;  
# Select TMP with mouse;  
(sele)>A>rename object: lig;  
lig>S>sticks;  
4AVL>H>lines;  
Command: isomesh lig_map, 2biw.map, 1.0, lig carve=1.6  
# click 4avl.map to hide the overall map.
```

Fifth step: Optimize the picture

```
center lig;  
lig_map>C>gray50;  
Command:  
>bg_color white  
>set mesh_width, 0.5  
>set ray_trace_fog, 0  
>set depth_cue, 0  
>set ray_shadows, off
```



Making movies: Rotating Molecule

Command:

```
mset 1, 180 %make a movie that has 180 frames;  
util.mroll 1, 180, 1 %make a movie that start at the  
1st frame and stop at the 180th frame with each  
frames rotating 1 degree;  
set ray_trace_frames, 1 %turn on the ray trace, it  
would be perfect!  
set cache_frames, 0 %turn out the cache frames
```

```
>save>movie>mpng
```



Making movies: Scene Translation

Making different states

State 1

```
>Scene>Store>F1;
```

State 2

```
>Scene>Store>F2;
```

State 3

```
>Scene>Store>F3;
```

.....

You can check your scene by

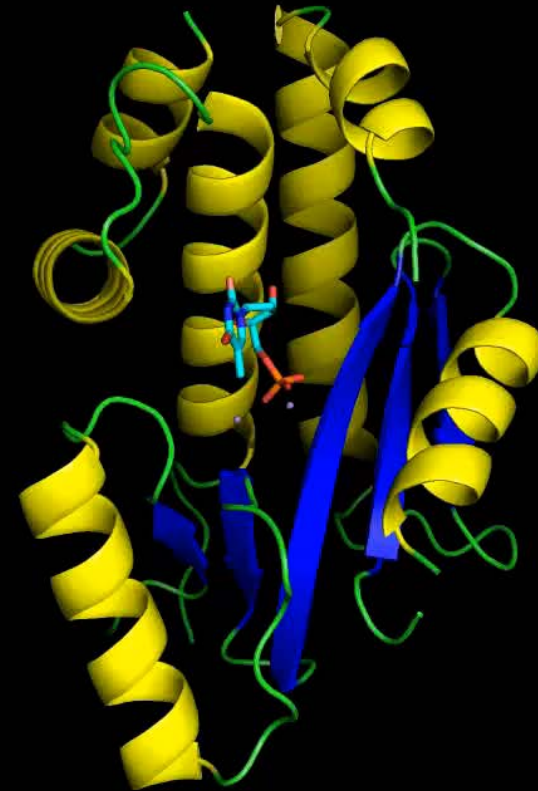
```
>Scene> Recall>Fn
```

Making movie

```
>Movies>Program>Scene Loop>....
```

```
% define the time and the translation mode;
```

```
>save>movie>mpng.
```



My Notes for using PyMol

- You will find the importance of “(sele)>A>copy to object” !!
- You will find it very very important to save a PyMol section!!!
- Rename the default generated sele_polar_conts term.

Thank You!!!

And Good Luck to Use PyMol!